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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
10/554,122	10/554,122 09/11/2006 Brenda M. Ogle		UM-30944/US-2/PCT	4639
72960 Casimir Jones, S	7590 06/14/2010 S.C. G WAY, SUITE 310 , WI 53562		EXAMINER	
2275 DEMING			STRZELECKA, TERESA E	
MIDDLETON,			ART UNIT	PAPER NUMBER
			1637	
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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary		Appli	cation No.	Applicant(s)				
		10/55	54,122	OGLE ET AL.				
		Exam	iner	Art Unit				
		TERE	SA E. STRZELECKA	1637				
The MAILING DATE of this communication appears on the cover sheet with the correspondence address Period for Reply								
A SHORTI WHICHEV - Extensions after SIX (6) - If NO period - Failure to re Any reply re	ENED STATUTORY PERIOD FOR IS LONGER, FROM THE M. of time may be available under the provisions MONTHS from the mailing date of this common life the provision of the maximum staply within the set or extended period for reply ceived by the Office later than three months a nt term adjustment. See 37 CFR 1.704(b).	AILING DATE OF of 37 CFR 1.136(a). In a unication. tutory period will apply a will, by statute, cause the	F THIS COMMUNICATION The event, however, may a reply be tire and will expire SIX (6) MONTHS from The application to become ABANDONE	N. nely filed the mailing date of this of (35 U.S.C. § 133).	•			
Status								
2a)∏ This 3)∏ Sinc	ponsive to communication(s) file action is <b>FINAL</b> .  e this application is in condition and the practice of th	th)⊠ This action for allowance exc	is non-final. cept for formal matters, pro		e merits is			
Disposition o	f Claims							
4a) C 5)	m(s) <u>1-10,13-17,51 and 52</u> is/are  Of the above claim(s) <u>9,10,16 and</u> m(s) is/are allowed.  m(s) <u>1-8,13-15,51 and 52</u> is/are  m(s) is/are objected to.  m(s) are subject to restrice	<u>d 17</u> is/are withdr rejected.	rawn from consideration.					
Application P	apers							
10)∏ The o	specification is objected to by the drawing(s) filed on is/are: icant may not request that any objected to accement drawing sheet(s) including that or declaration is objected to	a) ☐ accepted c ction to the drawing the correction is re	(s) be held in abeyance. Se quired if the drawing(s) is ob	e 37 CFR 1.85(a). jected to. See 37 C	` '			
Priority unde	r 35 U.S.C. § 119							
<ul> <li>12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).</li> <li>a) All b) Some * c) None of:</li> <li>1. Certified copies of the priority documents have been received.</li> <li>2. Certified copies of the priority documents have been received in Application No</li> <li>3. Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).</li> <li>* See the attached detailed Office action for a list of the certified copies not received.</li> </ul>								
Attachment(s)	ofgrange Cited (DTO 900)		4) 🗖 Intonia O	(PTO 442)				
2) Notice of D 3) Information	eferences Cited (PTO-892) raftsperson's Patent Drawing Review (P Disclosure Statement(s) (PTO/SB/08) )/Mail Date	TO-948)	4) Interview Summary Paper No(s)/Mail D 5) Notice of Informal F 6) Other:	ate				

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## **DETAILED ACTION**

1. This office action is in response to an amendment filed March 12, 2010. Claims 1-10, 13-17, 51 and 52 were previously pending, with claims 9, 10, 16 and 17 withdrawn from consideration.

Applicants did not amend any claims.

- 2. The declaration of Jeffrey Platt, MD filed on March 12, 2010 under 37 CFR 1.131 is sufficient to overcome the Gehrmann et al. reference. Consequently, all of the previously presented rejections are withdrawn.
- 3. Applicants' arguments regarding the priority date of the instant claims were considered persuasive, therefore it is considered to be April 24, 2003, the filing date of the provisional application No. 60/464,981.
- 4. This office action is made non-final because of new grounds for rejection.

## Claim Rejections - 35 USC § 103

- 5. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:
  - (a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.
- 6. This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

- 7. Claims 1-8, 13-15, 51 and 52 are rejected under 35 U.S.C. 103(a) as being unpatentable over Arstilla et al. (Science, vol. 286, pp. 958-961, 1999; cited in the IDS), Wagner et al. (PNAS USA, vol. 95, pp. 14447-14452, 1998; cited in the IDS and in the previous office action), Lebed et al. (J. Biomol. Struct. Dynam., vol. 18, pp. 813-823, 2001), Kamb et al. (U.S. Patent No. 6,060,240 A; issued May 2000), Cho et al. (Appl. Env. Microbiol., vol. 68, pp. 1425-1430, March 2002; cited in the previous office action) and Piechocki et al. (J. Immunol. Meth., vol. 259, pp. 33-42, January 2002).
- A) Regarding claims 1 and 13-15, Arstilla et al. teach determining lymphocyte diversity in a subject by obtaining cDNA from T-cells of a subject and amplifying the cDNA encoding CDR3 region of the TCR  $\beta$  receptor with V $\beta$ -, J $\beta$  and C $\beta$ -specific primers, followed by gel separation of amplified products and sequencing (page 958; page 959, first and third and fourth paragraphs; Fig. 1 and 2).

Regarding claims 1 and 13-15, Wagner et al. teach determining lymphocyte diversity in rheumatoid arthritis patients by amplification of cDNA encoding CDR3 region of the TCR  $\beta$  receptor from CD4 T-cells with V $\beta$ - and j $\beta$ -specific primers, followed by cloning and sequencing or hybridization to TCR N-D-N probes (page 14448, paragraphs 2-3 and 8; page 14449, first and second paragraph).

- B) Neither Arstilla et al. nor Wagner et al. teach hybridization of nucleic acids derived from lymphocytes to random nucleic acid molecules in order to determine lymphocyte diversity.
- C) Regarding claims 1, 2 and 13-15, Lebed et al. teach application of random oligonucleotide arrays to the determination of the CDR3 regions diversity in lymphocytes (page 813, last paragraph; page 814; page 815, paragraphs 1-3).

Regarding claim 3, Lebed et al. teach a chip (page 814, second paragraph).

Regarding claim 6, Lebed et al. teach random hexamers immobilized in different parts of the chip (page 814, second paragraph).

Regarding claim 7, Lebed et al. teach nucleic acids labeled with fluorophores (page 815, first paragraph).

Regarding claim 52, Lebed et al. teach labeled DNA molecules (page 815, first paragraph).

- D) Regarding claim 1, Kamb et al. teach a method comprising:
- a) providing:
- i) labeled nucleic acid molecules from a sample (col. 6, lines 7-11 and 28-38; col. 17, lines 29-42),
- ii) a population of nucleic acid molecules, wherein said population of nucleic acid molecules comprises random nucleic acid molecules (Fig. 6; col. 14, lines 23-64; col. 24, lines 58-62);
- b) hybridizing said labeled nucleic acid molecules or fragments of said labeled nucleic acid molecules with said population nucleic acid molecules (col. 6, lines 5-11 and 29-35);
- c) assessing hybridization of said labeled RNA nucleic acid molecules with said population of nucleic acid molecules to determine the frequency of hybridization (col. 6, lines 35-38), and
- d) quantifying the amount of hybridized nucleic acid (col. 6, lines 35-38; col. 24, lines 58-67; col. 25, lines 1-28).

Regarding claims 2-4, Kamb et al. teach beads (col. 5, lines 63-67; col. 6, lines 1-7; col. 9, lines 31-48; col. 10, lines 36-54).

Regarding claim 5, Kamb et al. teach flow cytometry (col. 6, lines 10, 11; col. 21, lines 25-39).

Regarding claim 7, Kamb et al. teach fluorophore labels (col. 6, lines 8, 9, 31, 32; col. 17, lines 29-42; col. 21, lines 25-39).

Regarding claim 8, Kamb et al. teach phycoerythrin (col. 21, line 64).

Regarding claims 51 and 52, Kamb et al. teach labeling mRNA or cDNA (col. 6, lines 30-32).

It would have been *prima facie* obvious to one of ordinary skill in the art at the time of the invention to have used solid support-bound random oligonucleotdes of Lebed et al. or Kamb et al. in the methods of detecting T-cell diversity of Arstilla et al. and Wagner et al. The motivation to do so is provided by Kamb et al. (col. 5, lines 43-47):

"The methods of the invention also provide other advantages, such as increasing the throughput of probes, boosting the generation of valuable data, and significantly lowering the time and cost of analysis."

Further, considering the fact that the estimated TCR diversity of Arstilla et al. was of the order of 10<sup>6</sup> (Table 1), using the random 15mer probes of Kamb et al. would provide 4<sup>15</sup>, or 1.1 x 10<sup>9</sup> different capture probes (col. 24, lines 62-64), allowing in principle capture of every single TCR variant present in a given cell type. Further, considering that FACS machines sort beads with a rate of about 100 million per hour (col. 21, lines 32-33), analysis of a multitude of different samples can be performed rapidly.

The motivation to do so is also provided by Wagner et al. (page 14452, last paragraph):

"Regardless of the precise mechanism for the loss of T cell diversity, these aberrations have important implications for the disease process and the way it is treated and studied. The fact that large proportions of the TCR repertoire are altered cannot remain without consequences for immunoresponsiveness. It is possible that the repertoire contraction will generate holes in the repertoire and therefore will lead to defective immune responses to selected antigens. The design of therapeutic approaches should consider that the RA repertoire already has lost diversity. So far, it

has been assumed that it would be beneficial to deplete T cells. If these patients have difficulties repopulating the T cell compartment and have to generate new T cells through self-replication, T cell-directed therapies will compromise further their ability to maintain diversity. It is, therefore, not surprising that treatment trials using T cell depletion were not successful and had substantial side effects (31, 41). Very different therapeutic approaches will have to be taken to correct repertoire aberrations in an attempt to control the disease process and its complications."

Therefore, it would have been *prima facie* obvious to one of ordinary skill in the art at the time of the invention that as precise as possible determination of lymphocyte diversity enabled effective diagnosis and treatment of infections and immune diseases.

E) None of the above references teaches quantitation of the amount of hybridized nucleic acids based on standard curves.

However, quantitation of either microarray data or flow cytometry date was known in the art of the invention, as evidenced by Cho et al. and Piechocki et al.

Cho et al. teach determining a number of expressed genes from array hybridization using standard curves obtained by hybridizing samples with known numbers of gene copies to an array of oligonucleotides, and creating a standard curve based on the measurements (page 1425, second paragraph; Fig. 1, 2; page 1426, last paragraph; page 1427; Fig. 3).

Piechocki et al. teach quantitation of anti-ErbB2 antibodies by flow cytometry based on standard curve (Abstract; page 35, first paragraph; page 38, last paragraph; page 39, first paragraph; Fig. 5).

Therefore it would have been *prima facie* obvious to one of ordinary skill in the art at the time of the invention to perform quantitative measurements of the T-cell diversity in the method of

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Arstilla et al., Wagner et al., Lebed et al. and Kamb et al., since the diversity was measured as the

total number of different sequences present in samples.

8. No claims are allowed.

Conclusion

Any inquiry concerning this communication or earlier communications from the examiner

should be directed to TERESA E. STRZELECKA whose telephone number is (571)272-0789. The

examiner can normally be reached on M-F (8:30-5:30).

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor,

Gary Benzion can be reached on (571) 272-0782. The fax phone number for the organization where

this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent

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system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

Teresa E Strzelecka Primary Examiner

Art Unit 1637

/Teresa E Strzelecka/

Primary Examiner, Art Unit 1637

June 10, 2010